

Short Communication

Identifying invertebrate invasions using morphological and molecular analyses: North American *Daphnia 'pulex'* in New Zealand fresh waters

Ian C. Duggan¹*, Karen V. Robinson², Carolyn W. Burns³, Jonathan C. Banks¹ and Ian D. Hogg¹

1 Centre for Biodiversity and Ecology Research, Department of Biological Sciences, University of Waikato, Private Bag 3105,

Hamilton, New Zealand

2 NIWA, Christchurch, P.O. Box 8602 Riccarton, Christchurch, New Zealand

3 Department of Zoology, University of Otago, P.O. Box 56, Dunedin, New Zealand

E-mail: i.duggan@waikato.ac.nz (ICD), karen.robinson@niwa.co.nz (KVR), carolyn.burns@otago.ac.nz (CWB), jonathan.banks@cawthron.org.nz (JCB), i.hogg@waikato.ac.nz (IDH)

*Corresponding author

Received: 24 February 2012 / Accepted: 13 June 2012 / Published online: 18 June 2012

Abstract

We used a DNA barcoding approach to identify specimens of the *Daphnia pulex* complex occurring in New Zealand lakes, documenting the establishment of non-indigenous North American *Daphnia 'pulex'*. Morphological delineation of species in this complex is problematic due to a lack of good morphological traits to distinguish the species, as there is a relatively high degree of morphological stasis within the group through evolutionary time. Accordingly, genetic analyses were used to determine the specific identity and likely geographic origin of this species. Morphologically, individuals most closely resembled *Daphnia pulicaria* or *Daphnia pulex* sensu lato, which cannot be separated morphologically. Furthermore, each of these taxa comprises separate species in North America and Europe, despite carrying the same names. We identified individuals using a 658 bp nucleotide portion of the mitochondrial cytochrome c oxidase subunit 1 gene (COI) as North American *Daphnia 'pulex'*, being distinct from European *Daphnia pulex* sensu stricto and *D. pulicaria*. North American *Daphnia 'pulex'* in New Zealand were first recorded in New Zealand from South Island lakes that are popular for overseas recreational fishers, indicating a possible source of introduction for this species (e.g. on/in fishing gear). Our study provides an additional example of how genetic techniques can be used for the accurate identification of non-indigenous taxa, particularly when morphological species determination is not possible. The growth of global databases such as GenBank and Barcode of Life Datasystems (BOLD) will further enhance this identification capacity.

Key words: biological invasions; biosecurity; cladocera, DNA barcoding

Introduction

The widespread establishment of non-indigenous species is progressively homogenising the world's floras and faunas. For example, the number of invertebrate invaders recognised in fresh waters in New Zealand has increased markedly in recent years, comprising species from diverse biogeographical realms (Duggan 2002; Duggan et al. 2006; Collier et al. 2011). Whereas some of these newly recognised invaders, such as the snail Melanoides tuberculata, are likely to have been established for some time (Duggan 2002), others such as the cladoceran Daphnia galeata and the calanoid copepod Skistodiaptomus pallidus represent recent invasions (Duggan et al. 2006), indicating that both the detection and invasion rates are increasing.

Here, we report on a new record of a *Daphnia* species belonging to the D. pulex species complex in New Zealand. The identification of species in this complex is difficult. Determination of species based on morphological methods is problematic due to a lack of reliable morphological traits as a result of morphological stasis combined with a poor taxonomy (Colbourne and Hebert 1996; Mergeay et al. 2008). Whereas the D. pulex complex was considered to comprise a small number of species that could be differentiated morphologically, a number of new species have been identified with the aid of genetic techniques (e.g., Hebert and Finston 1996; Kořinek and Hebert 1996). Although a number of the species within the *pulex* complex can be separated by morphological means, based on distinctive features that were identified following genetic

analyses, definitive identification of some members still requires genetic analysis (Benzie 2005). For example, the species Daphnia pulex and D. pulicaria, within the D. pulex complex, are currently impossible to distinguish confidently using morphological characters, but are genetically distinct (although see Mergeay 2008 and Cristescu 2012). Both Hebert (1995) and Benzie (2005) note that whereas there are some morphological features that are indicative of identity, definitive taxonomic assignment of these species depends on genetic determination. Furthermore, both Daphnia pulex and D. pulicaria sensu lato were thought to be widely distributed in the Northern Hemisphere, but have now been found to be genetically distinct (yet morphologically indistinguishable) between Europe and North America. Accordingly, the D. pulex complex is comprised of different species on each continent, although the same names persist in each respective location (Colbourne et al. 1998; Mergeay et al. 2008). An additional complicating factor is that, similar to several other Daphnia species, North American Daphnia 'pulex' and D. pulicaria are able to form viable hybrid populations, requiring allozyme analysis to confidently determine their status (e.g., Hebert et al. 1989). For example, based on DNA sequencing a North American Daphnia 'pulex' invasion was recognised in African lakes (Mergeay et al. 2005), although this species was later found by allozyme analysis to be a hybrid North American Daphnia 'pulex' × D. pulicaria (Mergeay et al. 2006).

DNA barcoding and other genetic techniques are now commonly used to confirm the morphological identifications of aquatic invaders, including in New Zealand (e.g., Stevens et al. 2002; Makino et al. 2010; Collier et al. 2011). Here we provide an example that required genetic analyses to identify a non-indigenous species that is a member of a species complex, and to determine its geographic origin.

Methods

To date we have observed the non-indigenous *Daphnia* species from a number of South Island lakes, where it is widespread (Table 1). The earliest samples we have of this species were collected from the Central Otago lakes, in March 2005. Morphologically, the *Daphnia* species conforms to *D. pulex* or *D. pulicaria* sensu lato, within the *D. pulex* complex, based on the keys of Hebert (1995) and Benzie (2005), and thus

586

requires genetic analyses for species level determination. The identity of the Daphnia species was resolved by direct sequencing of the polymerase chain reaction product from a 658 bp nucleotide portion of the mitochondrial cytochrome c oxidase subunit 1 gene (COI) from seven individuals from four lakes (three specimens from Lake Benmore, two from Lake Wanaka, and one each from Lake Hayes and Lake Heron) using the methods of Penton et al. (2004). The COI sequences we obtained were aligned with Daphnia pulicaria and D. pulex COI sequences downloaded from Genbank using the default values in Clustal X (Thompson et al. 1997). Alignment was straightforward as there were no insertions or deletions. Daphnia neoobtusa was used as an outgroup to root the phylogeny. Phylogenetic trees were estimated using the maximum likelihood GTR + G model as recommended by Jmodeltest (Posada 2008) and 1000 bootstrap replicates (Felsenstein 1985) were generated to assess support for the nodes in Mega (Tamura et al. 2011). All sequences were deposited in the project "Daphnia pulex in New Zealand" (NZPLB) in the Barcode of Life Datasystem (BOLD) database (http://www.barco dinglife.org) and cross referenced to GenBank (accession numbers HM622590 - HM622593, and JX150976 – JX150976). However, as mitochondrial genes are typically passed through the female lineage, and in North America the two species in question may hybridise as D. 'pulex' \times D. pulicaria (i.e., with maternal D. 'pulex'), we tested for the presence of hybrids using an allozyme analysis. Specifically, we assessed using cellulose allozyme electrophoresis (Hebert and Beaton 1989), whether individuals were homozygous (non-hybrid) or heterozygous (hybrid) at the lactate dehydrogenase locus (LDH: EC 1.1.1.27), an established method for determining hybridisation in these species, as per Hebert et al. (1989). Daphnia pulicaria is the only species that North American maternal D. 'pulex' is known to form hybrids with under natural conditions, and thus only this one allozyme locus was required for analysis (e.g., Hebert et al. 1989; Taylor and Hebert 1993). Identification of these species using LDH is commonly undertaken in North America; however, this is only possible if known D. 'pulex' and D. pulicaria are analysed with the unknown population, where relative mobilities can be assessed (slow = D. 'pulex' and fast = D. pulicaria). Unfortunately, this is not possible for situations where both species are not present

Table 1. South Island water bodies whe	re Daphnia 'pulex' h	has been sampled to date.
--	----------------------	---------------------------

Water Body	Geo-reference	Date of first record and Collector
Clutha River drainage system		
Lake Hayes	-44° 58' 48.00", 168° 48' 36.00"	3 Mar 05 (Burns)
Lake Johnson	-45° 00' 08.34", 168° 43' 53.88"	4 Mar 05 (Burns)
Lake Moke	-45° 00' 10.52", 168° 33' 51.34"	4 Mar 05 (Burns)
Lake Wakatipu	-45° 03' 28.80", 168° 39' 08.99"	5 Mar 05 (Burns)
Lake Wanaka	-44° 31' 33.19", 169° 06' 55.06"	10 Nov 09 (Burns)
Waitaki River drainage system		
Lake Benmore	-44° 22' 33.22", 170° 12' 50.14"	8 Dec 08 (Robinson)
Lake Waitaki	-44° 40' 29.56", 170° 23' 59.41"	10 Dec 08 (Robinson)
Lake Ruataniwha	-44° 16' 46.86", 170° 04' 09.49"	3 Jan 08 (Robinson)
Tekapo Canal	-44° 04' 14.93", 170° 21' 34.19"	5 Nov 07 (Robinson)
Ohau Canal	-44° 15' 39.37", 169° 58' 44.74"	5 Nov 07 (Robinson)
Canterbury rivers drainages		
Lake Heron	-43° 28' 36.62", 171° 10' 30.84"	17 Mar 09 (Robinson)
Lake Pegasus	-43° 18' 32.19", 172° 41' 48.85"	25 Mar 11 (Robinson)
Lake Sarah	-43° 02' 50.56", 171° 46' 36.55"	17 Mar 09 (Robinson)
Dunedin		
Ross Creek Reservoir	-45° 50' 48.93", 170° 29' 55.38"	20 Nov 08 (Burns)
Rossville Reservoir	-45° 48' 17.37", 170° 36' 01.97"	20 Nov 08 (Burns)
Sullivan's Dam	-45° 48' 26.03", 170° 31' 22.69"	22 Dec 08 (Burns)
Tomahawk Lagoon #2	-45° 54' 05.44", 170° 33' 02.18"	24 Aug 10 (Burns)
Southland		
Munro's Dam	-46° 29' 28.00", 168° 34' 53.00"	1 Oct 11 (Burns)
Lake Manapouri	-45° 33' 33.32", 167° 36' 24.25"	12 Mar 10 (Robinson)

(i.e., New Zealand). At least five individuals for allozyme analysis were analysed on fresh individuals collected from each of Lake Heron and Lake Wanaka.

Results

Based on morphological features (shape of head and rostrum, position of eye, spinulation of carapace margins, shape of postabdomen, relative size and setation of postabdominal processes, number and shape of postabdominal spines, size and number of teeth on combs of the postabdominal claw), adult female *Daphnia* collected most closely resembled *Daphnia pulicaria* sensu lato and *Daphnia pulex* sensu lato, which cannot be separated morphologically (Hebert 1995; Benzie 2005). Morphologically, the specimens collected from the Central Otago and Waitaki lakes differ from the descriptions of *D. pulex* in Benzie (2005) in having a more elliptical than oval carapace in lateral view and a tail spine longer than 0.3 carapace length, and a moderate-sized eye in a large optic vesicle that is set back from the frontal margin of the head. Specimens from Lake Wanaka kept in culture produced males and ephippia.

The phylogeny for our Daphnia specimens, estimated by maximum likelihood, indicate the New Zealand populations formed a monophyletic group with North American and African populations of Daphnia 'pulex' (Figure 1). Our sequences differed by 0.2 - 1.6% (1 to 10 nucleotides) from North American D. 'pulex' sequences in GenBank (for example, accession number DQ340843). Our sequences differed from GenBank sequences for Daphnia pulicaria by between 3% and 11% (between 16 to 62 nucleotides: GenBank accession numbers FJ427505 and EU152322). Our sequences differed from D. pulex sequences EU152320 and EU152321 in GenBank, described as D. pulex sensu stricto from Europe (Mergeay et al. 2008), by 11%, indicating that the South Island Daphnia originated from North America and not

Figure 1. Phylogeny hypothesis for Daphnia pulex and pulicaria sensu lato estimated using the maximum likelihood GTR + G model using COI sequences. Numbers to the left of nodes designate percentage bootstrap support for that node. The sequences from New Zealand Daphnia 'pulex' specimens are at the top of the figure.



Europe. Allozyme analyses showed that individuals were homozygous for the LDH locus, confirming the designation of North American D. '*pulex*' and not the hybrid D. *pulex* \times D. *pulicaria*.

Discussion

The phylogeny estimated for the Daphnia COI sequences placed our Daphnia with North American D. 'pulex', distinct from European Daphnia pulex sensu stricto and D. pulicaria from Europe or North America, with strong bootstrap support. Globally, non-indigenous populations of Daphnia species have been recorded from a number of locations outside their native ranges. Most notable is an African strain of Daphnia lumholtzi first recorded from a reservoir in Texas in 1991, which has subsequently spread widely in North America (Havel et al. 1995); this species was unmistakable, morphologically, from North American congeners due to the invaders' possession of a large distinct head spine. At the other extreme, North American Daphnia 'pulex' × D. pulicaria have also invaded several African lakes, displacing

588

native strains of the morphologically indistinguishable Daphnia pulex (Mergeay et al. 2005; 2006). Our sequences closely grouped with this African invader (Figure 1), although allozyme analysis indicates the New Zealand invader is not a hybrid with D. pulicaria. Cultures maintained in the laboratory in our study produced both males and ephippia, suggesting the populations may reproduce by sexual reproduction (although see Hebert et al. 1989; Innes et al. 2000). To date, all nonindigenous populations of North American Daphnia 'pulex' (including their D. pulicaria hybrids) recorded from Europe and Africa have been found to be obligate parthenogens (e.g., Mergeay et al. 2006; Mergeay pers. comm).

The African D. 'pulex' \times D. pulicaria invasion is likely to have occurred through the stocking of largemouth bass (*Micropterus salmoides*) from the USA (Mergeay et al. 2006), while the Daphnia lumholtzi invasion into North America was also likely to have occurred via fish stocking of Nile Tilapia from Africa (Havel et al. 2000). The transportation vector for North American D. 'pulex' into New Zealand is not obvious, although the distribution of the first recorded populations may provide some indication. Our earliest samples were collected from lakes in the Clutha River and Waitaki River drainages, including Lakes Benmore, Waitaki and Wakatipu, which contain well established populations of non-indigenous brown trout (Salmo trutta), rainbow trout (Oncorhynchus mykiss) and some contain land-locked salmon (Oncorhynchus nerka, O. tshawytscha). However, despite introductions of the latter three species being from North America, releases of these to New Zealand are historic (late 19th and 20th centuries), and any fish used in recent stockings of these lakes will have originated from hatcheries within New Zealand (McDowall 1990). Nevertheless, the presence of these salmonids makes these lakes extremely popular for recreational fishing. including for many international tourists, indicating a possible means of introduction in association with fishing and boating equipment. Fishing equipment, for example felt-soled waders, is a likely vector for another recent invader to New Zealand, a diatom, Didymosphenia geminata, which is currently restricted to South Island rivers where fishing is popular (Kilroy et al. 2008). Fouling of fishing lines by the small crustaceans Bythotrephes longimanus and Cercopagis pengoi, or their diapausing stages, is thought to have aided their spread through North American lakes (e.g., Jacobs and MacIsaac 2007). North American and European Daphnia pulex have been recorded from ballast tanks of transoceanic ships in the North American Great Lakes (Briski et al. 2010; Figure 1). However, such vessels do not enter freshwater ports in New Zealand, making this transportation vector unlikely in this instance. New Zealand has only one recognised native Daphnia species, the relatively uncommon Daphnia carinata, while cladoceran diversity overall is also low (Chapman and Green 1987). As such, a lack of strong niche overlap with existing species may have facilitated the establishment of North American D. 'pulex' in New Zealand (e.g., Dzialowski 2010).

The establishment of non-indigenous *Daphnia*, such as the North American *D. 'pulex'*, might have large effects on lake and pond biota. For example, Vanni (1986) found that *Daphnia 'pulex'* greatly reduced the abundances of both phytoplankton and other zooplankton (particularly rotifers) when it was introduced to enclosures in a North American pond. Similarly, a recent invasion of *Daphnia galeata* to New Zealand coincided with reductions in chlorophyll *a*

concentration, increased Secchi transparency and a reduction in rotifer abundances in Lake Puketerini (Weavers Lake) following its establishment (Balvert et al. 2009). Such changes are likely to influence higher trophic levels also.

The rate of invasion of freshwater invertebrates appears to be increasing, and the recent establishment of North American Daphnia 'pulex' in New Zealand is yet another example of this trend. Overall, our results provide further evidence of how genetic techniques can play a major role in the identification of new nonindigenous species, particularly if these new taxa belong to species complexes. In addition, we show that genetic techniques can also provide valuable information on donor (i.e., source) regions of new invaders, and may thus provide clues on invasion pathways, which may be used to reduce invasion rates. Such methods will become more applicable as global databases, such as GenBank and BOLD (Ratnasingham and Hebert 2007), grow in size. However, care must be taken for a lack of distinction between sibling species, or erroneous naming of taxa, in these databases. For example, in GenBank three distinct species are called simply "Daphnia pulex" (Figure 1). As eradication of aquatic invertebrate species is impossible once they are established. assessments of transportation vectors from significant donor regions identified by such means can be investigated to determine management strategies for reducing further introductions.

Acknowledgements

This research was funded in part by FRST grants UOWX0501 and UOWX0505. We thank M.Knox and G.Collins for laboratory assistance, and J. Mergeay and an anonymous referee for their helpful comments on this manuscript.

References

- Balvert SF, Duggan IC, Hogg ID (2009) Zooplankton seasonal dynamics in a recently filled mine pit lake: the effect of nonindigenous *Daphnia* establishment. *Aquatic Ecology* 43: 403–413, http://dx.doi.org/10.1007/s10452-008-9165-z
- Benzie JAH (2005) Cladocera: the genus *Daphnia* (including *Daphniopsis*). Leiden, The Netherlands, Backhuys Publishers, 376 pp
- Briski E, Cristescu ME, Bailey SA, MacIsaac HJ (2010) Use of DNA barcoding to detect invertebrate invasive species from diapausing eggs. *Biological Invasions* 13: 1325–1340, http://dx.doi.org/10.1007/s10530-010-9892-7
- Chapman MA, Green JD (1987) Zooplankton ecology. In: Viner AB (ed), Inland Waters of New Zealand. Department of Scientific and Industrial Research, Wellington, New Zealand, pp 225–263

- Colbourne JK, Crease TJ, Weider LW, Hebert PDN, Dufresne F, Hobæk A (1998) Phylogenetics and evolution of a circumarctic species complex (Cladocera: *Daphnia pulex*). *Biological Journal of the Linnean Society* 65: 347–365
- Colbourne JK, Hebert PDN (1996) The systematics of North American Daphnia (Crustacea: Anomopoda): a molecular phylogenetic approach. Philosophical Transactions of the Royal Society of London B 351: 349–360, http://dx.doi.org/ 10.1098/rstb.1996.0028
- Collier KJ, Demetras NJ, Duggan IC, Johnston TM (2011) Wild record of an apple snail in the Waikato River, Hamilton, and their incidence in freshwater aquaria. *New Zealand Natural Sciences* 36: 1–9
- Cristescu ME, Constantin A, Bock DG, Caceres CE, Crease TJ (2012) Speciation with gene flow and the genetics of habitat transitions. *Molecular Ecology* 21: 1411–1422, http://dx.doi.org/10.1111/j.1365-294X.2011.05465.x
- Duggan IC (2002) First record of a wild population of the tropical snail Melanoides tuberculata in New Zealand. New Zealand Journal of Marine and Freshwater Research 36: 825–829, http://dx.doi.org/10.1080/00288330.2002.9517135
- Duggan IC, Green JD, Burger DF (2006) First New Zealand records of three non-indigenous zooplankton species: *Skistodiaptomus pallidus*, *Sinodiaptomus valkanovi* and *Daphnia dentifera*. New Zealand Journal of Marine and Freshwater Research 40: 561–569, http://dx.doi.org/10.1080/ 00288330.2006.9517445
- Dzialowski AR (2010) Experimental effect of consumer identity on the invasion success of a non-native cladoceran. *Hydrobiologia* 652: 139–148, http://dx.doi.org/10.1007/s10 750-010-0326-4
- Felsenstein J (1985) Confidence limits on phylogenies: an approach using the bootstrap. *Evolution* 39: 783–791, http://dx.doi.org/10.2307/2408678
- Havel JE, Mabee WR, Jones JR (1995) Invasion of the exotic cladoceran Daphnia lumholtzi into North American reservoirs. Canadian Journal of Fisheries and Aquatic Science 52: 151–160, http://dx.doi.org/10.1139/f95-015
- Havel JE, Colbourne J, Hebert PDN (2000) Reconstructing the history of intercontinental dispersal in *Daphnia lumholtzi* by use of genetic markers. *Limnology and Oceanography* 45: 1414–1419, http://dx.doi.org/10.4319/lo.2000.45.6.1414
- Hebert PDN, Ward RD, Weider LG (1995) Clonal-diversity patterns and breeding-system variation in *Daphnia pulex*, asexual-sexual complex. *Evolution* 42: 147–159, http://dx.doi.org/10.2307/2409123
- Hebert PDN (1995) The *Daphnia* of North America: an illustrated fauna. CD ROM, University of Guelph, Guelph, ON, Canada
- Hebert PDN, Beaton MJ (1989) Methodologies for Allozyme Analysis Using Cellulose Acetate Electrophoresis. Helena Laboratories, Beaumont, TX
- Hebert PDN, Beaton MJ, Schwartz SS, Stanton DJ (1989) Polyphyletic origins of asexuality in *Daphnia pulex*. I. Breeding system variation and levels of clonal diversity. *Evolution* 43: 1004–1015, http://dx.doi.org/10.2307/2409581
- Hebert PDN, Finston TL (1996) A taxonomic reevaluation of North American Daphnia (Crustacea: Cladocera). II. New species of the D. pulex group from the southcentral United States and Mexico. Canadian Journal of Zoology 74: 632– 653, http://dx.doi.org/10.1139/z96-073
- Jacobs M, MacIsaac HJ (2007) Fouling of fishing line by the waterflea Cercopagis pengoi: a mechanism of humanmediated dispersal? Hydrobiologia 583: 119–126, http://dx.doi.org/10.1007/s10750-006-0487-3
- Innes DJ, Fox CJ, Winsor GL (2000) Avoiding the cost of males in obligatory asexual *Daphnia pulex* (Leydig). *Proceedings* of the Royal Society B 267: 991–997, http://dx.doi.org/10. 1098/rspb.2000.1101

- Kilroy K, Snelder TH, Floerl O, Vieglais CC, Dey KL (2008) A rapid technique for assessing the suitability of areas for invasive species applied to New Zealand's rivers. *Diversity* and Distributions 14: 262–272, http://dx.doi.org/10.1111/j. 1472-4642.2007.00406.x
- Kořinek V, Hebert PDN (1996) A new species complex of Daphnia (Crustacea, Cladocera) from the Pacific Northwest of the United States. Canadian Journal of Zoology 74: 1379– 1393, http://dx.doi.org/10.1139/z96-153
- Makino W, Knox MA, Duggan IC (2010) Invasion, genetic variation and species identity of the calanoid copepod *Sinodiaptomus valkanovi*. Freshwater Biology 55: 375– 386, http://dx.doi.org/10.1111/j.1365-2427.2009.02287.x
- McDowall RM (1990) New Zealand freshwater fish: a guide and natural history. Heinemann-Reed. Auckland, New Zealand, 553 pp
- Mergeay J, Aguilera X, Declerck S, Petrusek A, Huyse T, De Meester L (2008) The genetic legacy of polyploid Bolivian *Daphnia*: the tropical Andes as a source for the North and South American D. pulicaria complex. Molecular Ecology 17: 1789–1800, http://dx.doi.org/10.1111/j.1365-294X.2007. 03679.x
- Mergeay J, Verschuren D, De Meester L (2005) Cryptic invasion and dispersal of an American Daphnia in East Africa. Limnology and Oceanography 50: 1278–1283, http://dx.doi. org/10.4319/lo.2005.50.4.1278
- Mergeay J, Verschuren D, De Meester L (2006) Invasion of an asexual American water flea clone throughout Africa and rapid displacement of a native sibling species. *Proceedings of the Royal Society Series* B 273: 2839–2844, http://dx.doi. org/10.1098/rspb.2006.3661
- Penton EH, Hebert PDN, Crease TJ (2004) Mitochondrial DNA variation in North American populations of *Daphnia* obtusa: continentalism or cryptic endemism. *Molecular Ecology* 13: 97–107, http://dx.doi.org/10.1046/j.1365-294X. 2003.02024.x
- Posada D (2008) jModelTest: Phylogenetic Model Averaging. Molecular Biology and Evolution 25: 1253–1256, http://dx.doi.org/10.1093/molbev/msn083
- Ratnasingham S, Hebert PDN (2007) BOLD: The barcode of life data system (www.barcodinglife.org). *Molecular Ecology Notes* 7: 355–364, http://dx.doi.org/10.1111/j.1471-8286.2007. 01678.x
- Stevens MI, Hogg ID, Chapman MA (2002) The corophid amphipods of Tauranga Harbour, New Zealand: evidence of an Australian crustacean invader. *Hydrobiologia* 474: 147– 154, http://dx.doi.org/10.1023/A:1016575519015
- Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S (2011) MEGA5: Molecular Evolutionary Genetics Analysis Using Maximum Likelihood, Evolutionary Distance, and Maximum Parsimony Methods. *Molecular Biology and Evolution* 28: 2731–2739, http://dx.doi.org/10.1093/molbev/ msr121
- Taylor DJ, Hebert PDN (1993) Habitat-dependent hybrid parentage and differential introgression between neighboringly sympatric Daphnia species. Proceedings of the National Academy of Sciences 90: 7079–7083, http://dx.doi.org/10.1073/pnas.90.15.7079
- Thompson JD, Gibson TJ, Plewniak F, Jeanmougin F, Higgins DG (1997) The Clustal X windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acids Research* 24: 4876–4882, http://dx.doi.org/10.1093/nar/25.24.4876
- Vanni MJ (1986) Competition in zooplankton communities: suppression of small species by *Daphnia pulex*. *Limnology* and Oceanography 31: 1039–1056, http://dx.doi.org/10. 4319/10.1986.31.5.1039